

Spin-labelled glycolipid analogues: D-glucose series

Jean M.J. Tronchet ^{a,*}, Martina Zsély ^a, Michel Geoffroy ^b

^a *Institute of Pharmaceutical Chemistry, University of Geneva, Sciences II, CH-1211 Geneva 4, Switzerland*

^b *Department of Physical Chemistry, University of Geneva, Sciences II, CH-1211 Geneva 4, Switzerland*

Received 14 December 1994; accepted 29 March 1995

Abstract

Neoglycolipids bearing a paramagnetic probe in their lipophilic aglycon have been prepared. All belong to the D-glucose series, both anomers for the glucoside representatives, respectively β and α anomers in the *S*- and *C*-glucosyl series. Two different types of radical sites have been used, a relatively short-lived imino *N*-oxyl group for glucosides and a more stable *N*-acylamino *N*-oxyl moiety in the other cases. EPR spectra of these radical species afforded information on the conformation of the lipophilic chain in the vicinity of the paramagnetic probe.

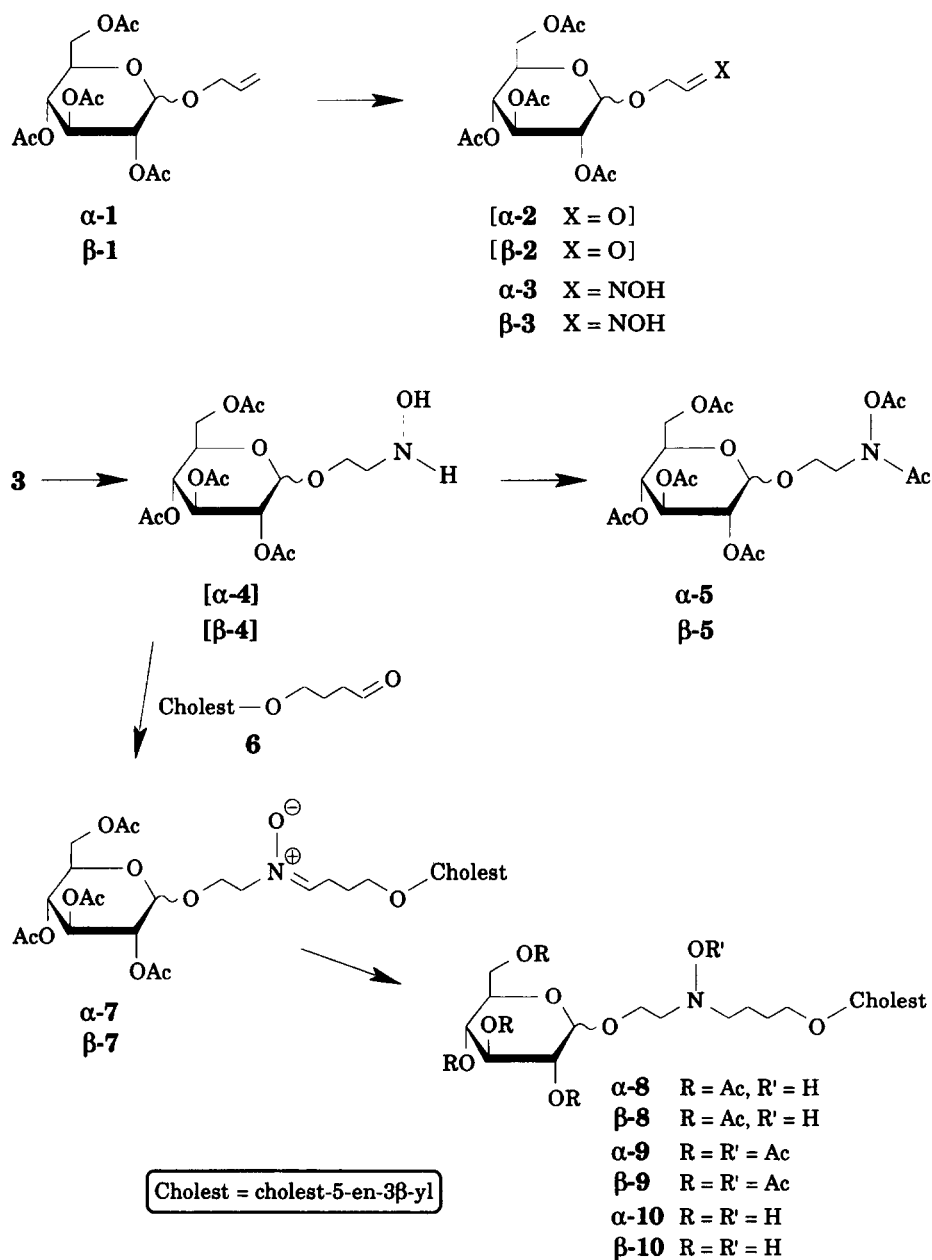
Keywords: EPR spectroscopy; Neoglycolipids; Spin labels; Glycosides

1. Introduction

Artificial glycolipids exhibit a large spectrum of useful properties: liquid crystals [1], specific detergents [2,3], immunomodulators [4], recognition markers for targeting drug delivery systems [5], or antiviral agents [6,7]. In most of these applications, the usefulness of neoglycolipids should be further increased by spin-labelling which would allow the monitoring of the interaction of these compounds with neighbouring molecules. We have described a number of close analogues of carbohydrate derivatives in which an oxygen atom is replaced with an NOH group. These compounds are spontaneously oxidized into the corresponding aminoxyl free radicals (for a review, see ref. [8]). These free radicals, the structurally closest possible paramagnetic analogues of natural sugars, have a half-life sufficient to allow good EPR spectra to be obtained but a little too short to afford comfortable monitoring of these compounds in biological media. For this reason, we also prepared sugar *N*-acylaminoxyls, markedly more stable than simple

* Corresponding author.

aminoxyls. We describe here *O*-, *S*-, and *C*-glucosyl compounds bearing a mono- or bi-antennary lipophilic chain, the monoantennary ones being spin-labelled. Part of this work has been reported at a Carbohydrate symposium [9].



Scheme 1.

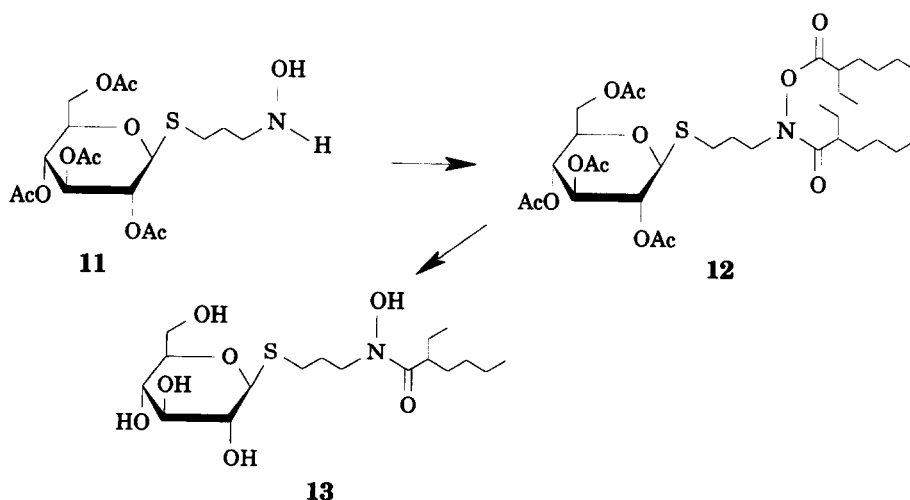
2. Results and discussion

Glycosidation of D-glucose with allyl alcohol in the presence of boron trifluoride etherate followed by peracetylation gave a resolvable 8:1 mixture (64%) of α -1 [10] and β -1 [10,11]. Each separated anomer was converted into the aldehyde **2**, which was not isolated in an analytically pure form but characterized by IR [$\nu_{C=O}$ 1757 (α -2), 1751 cm^{-1} (β -2)] and ^1H NMR [HC=O at δ 9.73 (α -2) and 9.65 (β -2), respectively, as badly resolved triplets]. Oximes α -3 and β -3 were obtained in good yield as 4:5 and 11:7 *E:Z* mixtures respectively (Scheme 1).

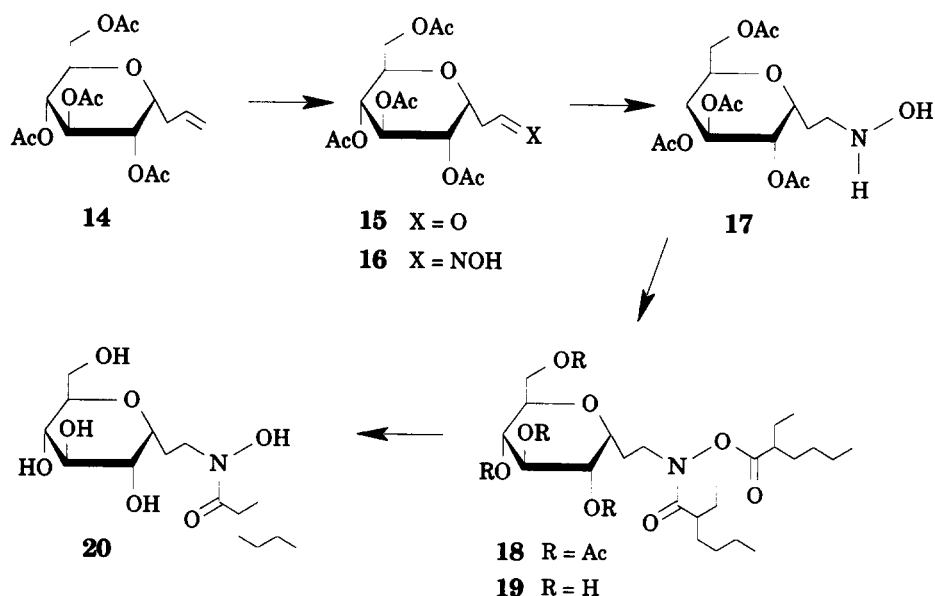
Upon sodium borohydride reduction in acidic medium [12,13], α -3 and β -3 afforded the corresponding hydroxylamines α -4 and β -4 in 63 and 69% yields, respectively. These compounds were converted in high yield into their *N,O*-diacetyl derivatives α -5 and β -5, thus confirming the structures of the parent compounds, particularly that of β -4 which was not obtained in a state of analytical purity.

The hydroxyamino group of compounds **4** was used as an anchor to fix a lipophilic chain onto the sugar moiety. Treated with the aldehyde **6** [14], each anomer of **4** yielded the expected nitrones α -7 (68%) and β -7 (64%) each as the *Z* diastereoisomer. These nitrones were characterized in particular by the appearance, in their NMR spectra, of a triplet (δ ca. 6.75, *J* 6 Hz) corresponding to the nitronic methine proton, thus establishing their *Z* configuration [15]. A high-yielding reduction (sodium borohydride) of α -7 and β -7 afforded α -8 and β -8, which were almost quantitatively *O*-acetylated to α -9 and β -9 or *O*-deacetylated to α -10 and β -10. The assignment of an α or β configuration to compounds **1**–**10** was straightforward. All these molecules adopted essentially the 4C_1 conformation, thus allowing an easy distinction between the β ($J_{1,2}$ 7–8 Hz) and the α anomers ($J_{1,2}$ 3.5–4 Hz).

Compounds such as **8** and **10**, in which a *N*-hydroxyimino group is included in an alkylidene chain, are spontaneously oxidized in the air to give the corresponding



Scheme 2.



Scheme 3.

aminoxyl, which is sufficiently long-lived to provide good EPR spectra, thus affording structural information concerning the isolated radical species in solution. Longer-lived radicals, necessary to study interactions between glycolipid analogues and macromolecules, were generated from hydroxamic acids. Such compounds have been prepared in the *S*- and *C*-glucosyl series.

The β -thioglucoside 11 [14], when treated with an excess of racemic 2-ethylhexanoyl chloride, underwent a di-*N,O*-acylation to the biantennary glycolipid analogue 12 in 77% yield (Scheme 2). Selective *O*-deacylation of 12 afforded 13 in 80% yield. Compounds 12 and 13 adopted an essentially pure 4C_1 conformation and their β configuration was confirmed by a large (9.5 Hz) $J_{1,2}$ coupling. The peracetylated *C*-glucosylpropene 14 [16] was oxidized (osmium tetroxide then sodium periodate) in 75% yield to give crystalline 15, which was converted into its crystalline *E*-oxime 16 (80%) (Scheme 3). Upon reduction, compound 16 gave the hydroxylamine 17 (67%), which was di-*O*-acylated to 18 (91%). *O*-Deacylation of 18 afforded a resolvable mixture of 19 (31%) and 20 (60%). The *C*-glucosyl compounds 15–20 also adopt principally a 4C_1 conformation and the $J_{1,2}$ couplings (4–5.5 Hz) of these α anomers are, as expected, a little larger than those of α -glucosides owing to the reduced electronegativity of their anomeric substituent. As a consequence of the presence of one or two (*RS*)-2-ethylhexanoyl groups on compounds 12, 13, and 18–20, these molecules are diastereoisomeric mixtures. However, due to the distance between the sugar moiety and the chiral center on the acyl group and the flexibility of the chain linking these two parts of the molecule, no difference between such diastereoisomers was revealed, either by chromatography or by NMR spectroscopy.

Diglyme solutions of hydroxylamines α -8 and β -8 and of hydroxamic acids 13 and

Table 1

EPR data (diglyme, a in G) of free radicals formed from selected sugar hydroxylamines and hydroxamic acids

Compound	Temp (°C)	g	a_N	a_H^β		a_H^γ	
α -8	110	2.0059	14.6	10.2	9.8	0.5	0.5
				10.2	9.8	0.5	0.5
β -8	110	2.0062	14.5	10.5	10.5	0.5	0.5
				10.5	8.2	0.5	0.5
13	70	2.0056	7.43	6.37	6.37		
20	70	2.0067	7.45	6.90	5.60		

20 were spontaneously oxidized to give the corresponding free radicals, EPR data of which are collected in Table 1. The spectra corresponding to α -8 and β -8 are very similar. The a_N hyperfine coupling constants correspond to the value expected from this type of compound [17]. The values of a_H^β of α -8 speak in favour of a local symmetry of the $\text{CH}_2\text{N}(\text{O}^\cdot)\text{CH}_2$ group and one can assign to each methylene group a pair of different values (10.2 and 9.8 G). Their sum corresponds to a situation often encountered [8,18], in which a fast exchange takes place between conformers where one C–H bond lies in the plane of the aminoxyl group. The situation is approximatively the same for β -8. In both cases, the number of long-range hyperfine couplings (a_H^γ) confirm the existence of a $(\text{CH}_2)_2\text{N}(\text{O}^\cdot)(\text{CH}_2)_2$ moiety.

Owing to the spin delocalization over the carbonyl group, free radicals derived from **13** and **20** exhibit hyperfine couplings approximatively halved relative to those from **8**. Taking this fact into account, the EPR data are close to those expected from a conformational equilibrium between forms in which one C–H bond of the methylene group would reside in the amidoxyl plane [8].

The biological usefulness of these compounds is being investigated.

3. Experimental

General.—The 2-ethylhexanoyl chloride was prepared from the corresponding carboxylic acid by the thionyl chloride method [19]. Solutions were concentrated under diminished pressure at $< 40^\circ\text{C}$. Melting points are uncorrected and were obtained with a Mettler FP52 melting-point microscope. TLC was performed on Silica Gel HF₂₅₄ (Merck) with detection by UV light or with phosphomolybdic acid– H_2SO_4 [20]. Column chromatography was conducted on Silica Gel 60 (0.063–0.2 mm, Merck). Silica Gel 60 (0.04–0.2 mm, Merck) was used for flash column chromatography [21]. IR spectra were recorded with a Perkin–Elmer FT-1600 or 1310 spectrometer and UV spectra with a Unicam SP 800 spectrophotometer. NMR spectra were recorded at 20°C for solutions in CDCl_3 (internal Me_4Si) with a Bruker WP 200 SY spectrometer (^1H , 200 MHz; ^{13}C , 50.4 MHz; chemical shifts in ppm from Me_4Si , δ units). Optical rotations were measured on solutions in CHCl_3 with a Schmidt–Haensch polarimeter. Mass spectra (EIMS, 70 eV) were recorded on a VG-70-70E spectrometer. EPR spectra were recorded on a Varian E-9 spectrometer (X band, 100 kHz modulation) equipped

with a variable temperature device and a UV irradiation facility. The g values were measured by using a DPPH sample and the magnetic field was calibrated with an NMR marker. All hyperfine coupling constants were checked by simulating the corresponding EPR spectra using a PC program developed in this laboratory [22].

Allyl 2,3,4,6-tetra-O-acetyl- α -D-glucopyranoside (α -1).—To a suspension of D-glucose (2 g, 11 mmol) in anhydrous allyl alcohol (40 mL) was added BF_3 etherate (0.2 mL, 1.63 mmol), and the mixture was heated under reflux for 2 h. The clear solution was cooled, concentrated to a syrup, then, to remove some minor impurities, submitted to column chromatography (9:1 CH_2Cl_2 –MeOH) which yielded 1.7 g (69.5%) of an 8:1 α/β syrupy mixture. This mixture was peracetylated with Ac_2O (20 mL) in pyridine (50 mL). After the usual work-up, the anomers were separated by column chromatography (1:1 hexane–ether) to give α -1 (2.5 g, 83%) and β -1 (0.3 g, 10%). Compound α -1 had mp 56–58°C; R_f 0.13 (1:1 hexane–ether); $[\alpha]_D^{27} +136^\circ$ (c 1.9, CHCl_3); $\nu_{\text{max}}^{\text{KBr}}$ 2938 (CH), 1740 (C=O), and 1652 cm^{-1} (C=C). ^1H NMR (CDCl_3): δ 5.89 (dddd, 1 H, $J_{2',3'a}$ 17.5, $J_{2',3'b}$ 10.5, $J_{2',1'a}$ 6, $J_{2',1'b}$ 5 Hz, H-2'), 5.52 (t, 1 H, $J_{3,2}=J_{3,4}=10$ Hz, H-3), 5.32 (dq, 1 H, $J_{3'a,3'b}$ 1.5 Hz, H-3'a), 5.23 (dq, 1 H, H-3'b), 5.11 (d, 1 H, $J_{1,2}$ 4 Hz, H-1), 5.09 (t 1 H, $J_{4,5}$ 10 Hz, H-4), 4.90 (dd, 1 H, H-2), 4.28 (dd, 1 H, $J_{5,6b}$ 5 Hz, H-6b), 4.20 (ddt, 1 H, $J_{3'b,1'b}$ 1.5 Hz, H-1'b), 4.10 (dd, 1 H, $J_{5,6a}$ 2.5, $J_{6a,6b}$ 13 Hz, H-6a), 4.08 (m, 1 H, H-5), 4.02 (ddt, 1 H, $J_{1'a,1'b}$ 13, $J_{1'a,3'a}$ 1.5 Hz, H-1'a), 2.11, 2.07, 2.02, and 2.00 (4 s, each 3 H, 4 OAc). EIMS: m/z (%) 388 (0.1, M^{+}), 347 (0.2, $\text{M}^{+}-\text{CH}_2-\text{CH}=\text{CH}_2$), 331 (1.5, $\text{M}^{+}-\text{OAc}$), 200 (17.8), 169 (28.3), 157 (58), 115 (70.4), 98 (68.4), 81 (100), 69 (52), 55 (18.7), and 45 (9.5). Anal. Calcd for $\text{C}_{17}\text{H}_{24}\text{O}_{10}$ (388.37): C, 52.58; H, 6.23. Found: C, 52.39; H, 6.12.

Allyl 2,3,4,6-tetra-O-acetyl- β -D-glucopyranoside (β -1).—Obtained as described for α -1; mp 89–90°C (from ether); lit. mp 88°C [11], 86°C [10]; R_f 0.11 (1:1 hexane–ether); $[\alpha]_D^{25} -11.7^\circ$ (c 1.0, CHCl_3); $\nu_{\text{max}}^{\text{KBr}}$ 2950 (CH), 1754 (C=O), and 1650 cm^{-1} (C=C). ^1H NMR (CDCl_3): δ 5.84 (dddd, 1 H, $J_{2'a,3'a}$ 17.5, $J_{2',1'b}$ 4.5, $J_{2',1'a}$ 6.5, $J_{2',3'b}$ 10.5 Hz, H-2'), 5.26 (dq, 1 H, $J_{3'a,3'b}=J_{3'a,1'a}=J_{3'a,1'b}=1.5$ Hz, H-3'a), 5.20 (t, 1 H, $J_{3,2}=J_{3,4}=9.5$ Hz, H-3), 5.18 (dq, 1 H, H-3'b), 5.06 (t, 1 H, $J_{4,5}$ 9.5 Hz, H-4), 5.00 (dd, 1 H, $J_{2,1}$ 8 Hz, H-2), 4.54 (d, 1 H, H-1), 4.32 (ddt, 1 H, $J_{1'a,1'b}$ 13.2 Hz, H-1'b), 4.26 (dd, 1 H, $J_{6b,5}$ 5, $J_{6a,6b}$ 11.7 Hz, H-6b), 4.11 (dd, 1 H, $J_{6a,5}$ 2.5 Hz, H-6a), 4.08 (ddt, 1 H, H-1'a), 3.66 (ddd, 1 H, H-5), 2.07, 2.02, 2.00, and 1.99 (4 s, each 3 H, 4 OAc). EIMS: m/z (%) 347 (0.2, $\text{M}^{+}-\text{CH}_2-\text{CH}=\text{CH}_2$), 331 (0.9, $\text{M}^{+}-\text{OAc}$), 243 (8.6), 200 (15.3), 169 (17.2), 157 (38), 115 (55), 98 (62), 81 (100), 69 (51), 55 (13.6), and 45 (7.7). Anal. Calcd for $\text{C}_{17}\text{H}_{24}\text{O}_{10}$ (388.37): C, 52.58; H, 6.23. Found: C, 52.33; H, 6.38.

2-Oxoethyl 1,3,4,6-tetra-O-acetyl- α -D-glucopyranoside (α -2).—To a solution of α -1 (1.2 g, 3.09 mmol) in a mixture of 1,4-dioxane (20 mL) and water (2 mL) was added OsO_4 (15 mg, 0.059 mmol); after 40 min stirring, saturated aq NaIO_4 (1.5 g, 7.01 mmol) was slowly added to the mixture, which was further stirred for 1.5 h, filtered, and concentrated to a syrup. Column chromatography (1:1 hexane–EtOAc) yielded α -2 (0.82 g, 67.6%) as a somewhat unstable syrupy material which was not obtained in analytically pure form but used for the preparation of α -3; R_f 0.13 (1:1 hexane–EtOAc); $\nu_{\text{max}}^{\text{NaCl}}$ 2963 (CH) and 1757 cm^{-1} (C=O). ^1H NMR (CDCl_3): δ 9.73 (t, 1 H, H-2'), 5.53 (t, 1 H, H-3), 5.13 (d, 1 H, H-1), 5.10 (t, 1 H, H-4), 4.92 (dd, 1 H, H-2), 4.30 (d, 2 H, H-1'a,1'b), 4.35–4.05 (m, 3 H, H-6a,b + H-5), 2.12, 2.10, 2.06, and 2.05 (4 s, each 3 H,

4 OAc). EIMS: m/z (%) 349 (0.9, $M^{++} - \text{Ac}$), 331 (7.9, $M^{++} - \text{OAc}$), 243 (4.7), 169 (76.5), 157 (50.7), 115 (100), 109 (83.7), 98 (66), 21 (76), 69 (62), and 57 (18).

2-Oxoethyl 1,3,4,6-tetra-O-acetyl- β -D-glucopyranoside (β -2).—Obtained as described under α -2, yield 63%; R_f 0.14 (2:3 hexane–EtOAc); $\nu_{\text{max}}^{\text{NaCl}}$ 2960 (CH), and 1751 cm^{-1} (C=O). ^1H NMR (CDCl_3): δ 9.65 (t, 1 H, H-2'), 5.20 (t, 1 H, H-2), 5.10 (dt, 1 H, H-4), 4.60 (d, 1 H, H-1), 4.35–4.02 (m, 4 H, H-1'a,1'b + H-6a,b), 3.20 (ddd, 1 H, H-5), 2.10, 2.02, 2.00 (each s, 4×3 H, 4 OAc). EIMS: m/z (%) 389 (0.7, $M^{++} - 1$), 331 (23, $M^{++} - \text{OAc}$), 243 (17), 200 (22), 169 (93), 145 (62), 115 (100), 81 (83), 69 (61), and 61 (17).

(E)- and (Z)-2-(Hydroxyimino)ethyl 2,3,4,6-tetra-O-acetyl- α -D-glucopyranoside (α -3).—To a solution of α -2 (5.5 g, 14 mmol) in EtOH (50 mL) and pyridine (5 mL) was added hydroxylamine hydrochloride (2.1 g, 30 mmol). After 14 h stirring at 20°C the mixture was concentrated, diluted with CH_2Cl_2 (200 mL), washed (water), dried (Na_2SO_4), and submitted to column chromatography (2:3 hexane–EtOAc) to yield α -3 (5.28 g, 92.5%) as a 4:5 *E/Z* mixture, mp 138–139°C (from ether); R_f 0.28 (1:1 hexane–EtOAc); $[\alpha]_D^{27} + 128.1^\circ$ (c 1.6, CHCl_3); $\nu_{\text{max}}^{\text{KBr}}$ 3470 (OH), 2979 (CH), 1745 (C=O), and 1705 cm^{-1} (C=N). ^1H NMR (CDCl_3): δ 8.20 (bs, =N–OH *Z*), 8.05 (bs, =N–OH *E*), 7.50 (t, $J_{2',1'}$ 6 Hz, H-2' *E*), 6.95 (t, $J_{2',1'}$ 3.5 Hz, H-2' *Z*), 5.50 (t, 1 H, $J_{3,2} = J_{3,4} = 9.5$ Hz, H-3), 5.12 (d, 1 H, $J_{1,2}$ 4 Hz, H-1), 5.10 (t, 1 H, H-4), 4.90 (dd, 1 H, H-2), 4.12–3.98 (m, 5 H, H-1'a,1'b, H-6a,b, H-5), and 2.12–1.98 (each s, 4×3 H, 4 OAc). EIMS: m/z (%) 345 (4), 331 (11, $M^{++} - \text{OCH}_2\text{CH}=\text{N} \sim \text{OH}$), 243 (15), 200 (23), 157 (100), 115 (87), 92 (78), 81 (57), 74 (32), and 58 (10). Anal. Calcd for $\text{C}_{16}\text{H}_{23}\text{NO}_{11}$ (405.36): C, 47.41; H, 5.72; N, 3.46. Found: C, 47.39; H, 5.60; N, 3.40.

(E)- and (Z)-2-(Hydroxyimino)ethyl 2,3,4,6-tetra-O-acetyl- β -D-glucopyranoside (β -3).—Applied to β -2 the procedure described for the preparation of α -3 yielded β -3 (89%) as an 11:7 mixture of *E/Z* isomers; R_f 0.24 (2:3 hexane–EtOAc); mp 107–108°C (from ether); $[\alpha]_D^{19} - 14.44^\circ$ (c 1.4, CHCl_3); $\nu_{\text{max}}^{\text{KBr}}$ 3485 (N–OH), 2968 (CH), 1760 (C=O), and 1720 cm^{-1} (C=N). ^1H NMR (CDCl_3): δ 7.75 (bs, =N \sim OH *Z*), 7.58 (bs, =N \sim OH *E*), 7.49 (t, $J_{2',1'}$ 5 Hz, H-2' *E*), 6.92 (t, $J_{2',1'}$ 3 Hz, H-2' *Z*), 5.30–5.00 (m, 3 H, $J_{2,3} = J_{3,4} = 9.5$ Hz, H-2, H-3, H-4), 4.70–4.53 (m, ~ 2 H, H-1, H-1'a,1'b *Z*), 4.44–4.10 (m, ~ 3 H, H-1'a,1'b *E*, H-6a,b), 3.75 (m, 1 H, H-5), and 2.13–2.00 (each s, 4×3 H, $4 \times \text{OAc}$). EIMS: m/z (%) 375 (0.5), 345 (5, $M^{++} - \text{AcOH}$), 331 (7, $M^{++} - \text{OCH}_2\text{CH}=\text{N} \sim \text{OH}$), 286 (4), 243 (13), 200 (22), 157 (68), 115 (90), 98 (100), 81 (93), and 73 (59). Anal. Calcd for $\text{C}_{16}\text{H}_{23}\text{NO}_{11}$ (405.36): C, 47.41; H, 5.72; N, 3.46. Found: C, 47.37; H, 5.65; N, 3.47.

2-(Hydroxyamino)ethyl 2,3,4,6-tetra-O-acetyl- α -D-glucopyranoside (α -4).—To a solution of α -3 (1.0 g, 2.47 mmol) in MeOH (10 mL) was added NaBH_4 (1.7 g, 27 mmol), and the pH was kept at 2–3 by dropwise addition of 6 M methanolic HCl. After the pH had stabilized to 3 without further addition of acid, the mixture was stirred for another 2 h, then concentrated, and the residue dissolved in citrate/HCl Titrisol pH 3 buffer (20 mL). The aqueous phase was washed (CH_2Cl_2 , 5 mL), brought to pH 8–8.5 (aq 10% NaOH saturated with NaCl), and extracted with CH_2Cl_2 (3×20 mL). The combined organic phase was dried (Na_2SO_4) and subjected to column chromatography (19:1 CH_2Cl_2 –MeOH) to give syrupy α -4 (0.63 g, 63%); R_f 0.20 (19:1 CH_2Cl_2 –MeOH); $\nu_{\text{max}}^{\text{NaCl}}$ 3425 (NH), 3390 (OH), 2957 (CH), and 1747 cm^{-1} (C=O). ^1H NMR (CDCl_3): δ

6.20–5.50 (bs, 2 H, NH, N-OH), 5.45 (t, 1 H, $J_{3,2} = J_{3,4} = 9.5$ Hz, H-3), 5.12 (d, 1 H, $J_{1,2} = 3.5$ Hz, H-1), 5.10 (t, 1 H, $J_{4,5} = 9.5$ Hz, H-4), 4.88 (dd, 1 H, H-2), 4.00–4.35 (m, 3 H, H-6a,b + H-5), 3.90 and 3.65 (2 m, each 1 H, H-1'a,1'b), 3.12 (t, 2 H, H-2'a,b), 2.10, 2.08, 2.05, and 2.02 (4 s, each 3 H, 4 OAc). EIMS: m/z (%) 408 (1.3, $M^{++} + 1$), 390 (0.2, $M^{++} - OH$), 366 (0.8, $M^{++} - Ac$), 350 (0.25), 331 (1.8), 289 (1.1), 169 (62.6), 139 (11.3), 127 (32), 109 (100), 102 (67.5), and 60 (79.5). Anal. Calcd for $C_{16}H_{25}NO_{11}$ (407.38): C, 47.17; H, 6.19; N, 3.44. Found: C, 46.79; H, 6.28; N, 3.57.

2-(Hydroxyamino)ethyl 2,3,4,6-tetra-O-acetyl- β -D-glucopyranoside (β -4).—Treatment of β -3 as described for the preparation of α -4 yielded syrupy β -4 (69%) which was not obtained in an analytically pure form but characterized as its diacetyl derivative β -5; R_f 0.28 (19:1 CH_2Cl_2 –MeOH); ν_{max}^{NaCl} 3470 (NH), 3280 (OH), and 1754 cm^{-1} (C=O). 1H NMR ($CDCl_3$): δ 5.30–4.90 (bs, 2 H, NH, N-OH), 5.22 (t, 1 H, $J_{3,2} = J_{3,4} = 9.5$ Hz, H-3), 5.08 (t, 1 H, H-4), 5.00 (dd, 1 H, $J_{2,1} = 8$ Hz, H-2), 4.55 (d, 1 H, H-1), 4.22 (m, 2 H, $J_{5,6b} = 5$, $J_{5,6a} = 2.5$, $J_{6a,6b} = 12$ Hz, H-6a,b), 3.98 and 3.85 (2 m, each 1 H, H-1'a,1'b), 3.72 (ddd, 1 H, H-5), 3.08 (t, 2 H, H-2'a,b), 2.10, 2.08, 2.02, and 2.00 (4 s, each 3 H, 4-OAc). EIMS: m/z (%) 408 (0.06, $M^{++} + 1$), 392 (10.17, $M^{++} - OH$), 331 (0.7), 229 (1.9), 187 (2), 169 (38), 157 (6.7), 109 (43), 86 (100), 72 (16), and 60 (14.5).

2-(N-Acetoxyacetamido)ethyl 2,3,4,6-tetra-O-acetyl- α -D-glucopyranoside (α -5).—A solution of α -4 (0.2 g, 0.49 mmol) in a mixture of pyridine (5 mL) and Ac_2O (1 mL) was kept 14 h at room temperature. After the usual work-up, the residue was subjected to column chromatography (3:2 hexane–acetone) to give α -5 (0.19 g, 79%) as a syrup; R_f 0.11 (3:2 hexane–acetone); $[\alpha]_D^{23} + 45.2^\circ$ (c 1.0, $CHCl_3$); ν_{max}^{NaCl} 2956 (CH), 1795, 1743 (C=O, acetyl), and 1680 cm^{-1} (C=O, NAc). 1H NMR ($CDCl_3$): δ 5.45 (t, 1 H, $J_{3,2} = J_{3,4} = 9.5$ Hz, H-3), 5.05 (m, 2 H, $J_{1,2} = 3.5$ Hz, H-1 + H-4), 4.35 (dd, 1 H, H-2), 4.18 (m, 2 H, $J_{5,6b} = 5$, $J_{5,6a} = 3$, $J_{6a,6b} = 13$ Hz, H-6a,b), 4.10–3.50 (m, 5 H, H-1'a,b + H-2'a,b + H-5), 2.25 (s, 3 H, NAc), 2.12, 2.10, 2.05, and 2.02 (each s, 5 \times 3 H, 5 \times OAc). EIMS: m/z (%) 491 (0.1, M^{++}), 432 (0.2, $M^{++} - OAc$), 331 (4), 169 (64), 144 (55), 127 (16.5), 102 (100), 81 (29), 73 (11.6), and 60 (18.9). Anal. Calcd for $C_{20}H_{29}NO_{13}$ (491.45): C, 48.88; H, 5.95; N, 2.85. Found: C, 48.65; H, 6.01; N, 2.95.

2-(N-Acetoxyacetamido)ethyl 2,3,4,6-tetra-O-acetyl- β -D-glucopyranoside (β -5).—Treated as described for the preparation of α -5, β -4 gave β -5 (82%) as a white crystalline powder, mp 110–111°C (from ether); R_f 0.23 (3:2 hexane–acetone); $[\alpha]_D^{21} - 12.36^\circ$ (c 0.9, $CHCl_3$); ν_{max}^{KBr} 2880 (CH), 1789, 1759, 1744 (C=O, OAc), and 1688 cm^{-1} (C=O, NAc). 1H NMR ($CDCl_3$): δ 5.15 (t, 1 H, $J_{3,2} = J_{3,4} = 10$ Hz, H-3), 5.08 (t, 1 H, H-4), 5.00 (dd, 1 H, $J_{2,1} = 7.5$ Hz, H-2), 4.52 (d, 1 H, H-1), 4.18 (m, 2 H, H-6a,b), 4.05–3.60 (m, 5 H, H-1'a,b + H-2'a,b + H-5), 2.18 (s, 3 H, NAc), 2.10, 2.08, 2.05, 2.02, and 1.98 (5 s, each 3 H, 5 OAc). EIMS: m/z (%) 492 (1.1, $M^{++} + 1$), 389 (0.3, $M^{++} - AcOH - CH_3CO$), 331 (10.7), 169 (65), 144 (83.5), 127 (14.7), 109 (65.9), 102 (100), 81 (23.5), and 60 (10.8). Anal. Calcd for $C_{20}H_{29}NO_{13}$ (491.45): C, 48.88; H, 5.95; N, 2.85. Found: C, 48.90; H, 5.85; N, 2.90.

3-Aza-7-(cholest-5-en-3 β -yloxy)hept-3-enyl 2,3,4,6-tetra-O-acetyl- α -D-glucopyranoside N-oxide (α -7).—To a solution of α -5 (0.6 g, 1.47 mmol) in CH_2Cl_2 (20 mL) was added **6** (0.8 g, 1.75 mmol), and the mixture was stirred 2 h at room temperature. The concentrated mixture was then subjected to column chromatography (19:1 ether–MeOH)

to give α -7 (0.85 g, 68%); R_f 0.13 (19:1 ether–MeOH); $[\alpha]_D^{20} + 42.3^\circ$ (c 1.0, CHCl_3); $\lambda_{\text{max}}^{\text{EtOH}}$ 202 (ϵ 5160) and 234 nm (8998); $\nu_{\text{max}}^{\text{KBr}}$ 2937 (CH), 2868 (CH), and 1758 cm^{-1} (C=O). $^1\text{H NMR}$ (CDCl_3): δ 6.87 (t, 1 H, $J_{4',5'a,b}$ 6 Hz, H-4'), 5.39 (t, 1 H, $J_{3,2} = J_{3,4} = 10$ Hz, H-3), 5.32 (m, 1 H, =CH chol), 5.08 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 5.06 (t, 1 H, $J_{4,5}$ 10 Hz, H-4), 4.90 (dd, 1 H, H-2), 4.33–3.68 (ms, 7 H, H-6a,b, H-5, H-1'a,b, H-7'a,b), 3.50 (dt, 2 H, H-2'a,b), 3.13 (tt, 1 H, $J_{\text{Chol}2a,3} = J_{\text{Chol}4a,3} = 11$ Hz, $J_{\text{Chol}2b,3} = J_{\text{Chol}4b,3} = 4.5$ Hz, H-3 chol), 2.66–0.54 (m, chol skeleton), 2.57 (q, 2 H, H-5'a,b), 2.08, 2.06, 2.03, and 2.00 (4 s, each 3 H, 4 OAc). EIMS: m/z (%) 829 (1, $\text{M}^{+\cdot}$), 770 (2.5), 531 (4.5), 462 (20, $\text{M}^{+\cdot} - \text{Ochol}$), 386 (40, Ochol), 368 (90), 353 (38), 247 (40), 213 (38), 169 (82), 145 (76), 115 (80), 95 (78), and 55 (72). Anal. Calcd for $\text{C}_{47}\text{H}_{75}\text{NO}_{12}$ (846.12): C, 66.72; H, 8.93; N, 1.66. Found: C, 66.44; H, 8.88; N, 1.66.

3-Aza-7-(cholest-5-en-3 β -yloxy)hept-3-enyl 2,3,4,6-tetra-O-acetyl- β -D-glucopyranoside N-oxide (β -7).—Treatment of β -5 and 6 as described for the preparation of α -7 gave β -7 (64%) as a white foam; R_f 0.09 (19:1 ether–MeOH); $[\alpha]_D^{25} - 26.1^\circ$ (c 0.77, CHCl_3); $\lambda_{\text{max}}^{\text{EtOH}}$ 233 nm (ϵ 8472); $\nu_{\text{max}}^{\text{KBr}}$ 2936 (CH), 2867 (CH), and 1757 cm^{-1} (C=O). $^1\text{H NMR}$ (CDCl_3): δ 6.75 (t, 1 H, $J_{4',5'a,b}$ 6 Hz, H-4'), 5.33 (m, 1 H, =CH chol), 5.20 (t, 1 H, $J_{3,2} = J_{3,4} = 9.5$ Hz, H-3), 5.12 (t, 1 H, $J_{4,5}$ 9.5 Hz, H-4), 4.98 (dd, 1 H, $J_{2,1}$ 7.5 Hz, H-2), 4.53 (d, 1 H, H-1), 4.35–3.80 (ms, 6 H, H-6a,b + H-1'a,b + H-7'a,b), 3.72 (m, 1 H, H-5), 3.50 (t, 2 H, H-2'a,b), 3.15 (tt, 1 H, H-3 chol), 2.70–0.60 (m, chol skeleton), 2.60 (m, 2 H, H-5'a,b), 2.10, 2.05, and 2.03 (each s, 4×3 H, $4 \times \text{OAc}$). EIMS: m/z (%) 846 (0.5, $\text{M}^{+\cdot}$), 829 (4, $\text{M}^{+\cdot} - \text{OH}^\cdot$), 770 (10), 462 (100, $\text{M}^{+\cdot} - \text{Ochol}$), 386 (28, Ochol), 368 (70), 247 (22), 169 (100), 109 (65), 72 (67), and 55 (43). Anal. Calcd for $\text{C}_{47}\text{H}_{75}\text{NO}_{12}$ (846.12): C, 66.72; H, 8.93; N, 1.66. Found: C, 66.50; H, 8.93; N, 1.80.

3-Aza-7-(cholest-5-en-3 β -yloxy)-3-hydroxyheptyl 2,3,4,6-tetra-O-acetyl- α -D-glucopyranoside (α -8).—To a solution of α -7 (0.7 g, 0.83 mmol) in MeOH (20 mL) was added NaBH_3CN (0.2 g, 0.2 mmol), and the pH was kept at 3 as described for the preparation of α -4. After completion of the reaction (TLC), the mixture was neutralized (pH 7–8), concentrated, and extracted with CH_2Cl_2 (50 mL). The organic phase was washed (water, 10 mL), dried (Na_2SO_4), concentrated, and subjected to column chromatography (4:1 ether–petroleum ether) to give α -8 (0.59 g, 84%) as a white foam; R_f 0.16 (4:1 ether–petroleum ether); $[\alpha]_D^{21} + 48.2^\circ$ (c 0.9, CHCl_3); $\nu_{\text{max}}^{\text{KBr}}$ 3472 (OH), 2935 (CH), 2866 (CH), and 1753 cm^{-1} (C=O). $^1\text{H NMR}$ (CDCl_3): δ 5.50 (t, 2 H, $J_{3,2} = J_{3,4} = 10$ Hz, H-3 + N-OH), 5.33 (m, 1 H, =CH chol), 5.11 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 5.02 (t, 1 H, $J_{4,5}$ 10 Hz, H-4), 4.86 (dd, 1 H, H-2), 4.28 (m, 2 H, H-5 + H-6b), 4.10 (m, 1 H, H-6a), 3.90 (m, 1 H, H-1'b), 3.73 (m, 1 H, H-1'a), 3.48 (m, 2 H, H-7'a,b), 3.14 (tt, 1 H, H-3 chol), 2.85 (m, 2 H, H-2'a,b), 2.70 (m, 2 H, H-4'a,b), 2.45–0.60 (m, chol skeleton), 2.10, 2.08, 2.03, and 2.00 (4 s, each 3 H, 4 OAc). EIMS: m/z (%) 531 (0.1), 512 (0.4), 462 (12, $\text{M}^{+\cdot} - \text{Ochol}$), 368 (100, Ochol), 247 (50), 145 (68), 81 (80), and 60 (68). Anal. Calcd for $\text{C}_{47}\text{H}_{77}\text{NO}_{12}$ (848.14): C, 66.56; H, 9.15; N, 1.65. Found: C, 66.48; H, 9.23; N, 1.74.

3-Aza-7-(cholest-5-en-3 β -yloxy)-3-hydroxyheptyl 2,3,4,6-tetra-O-acetyl- β -D-glucopyranoside (β -8).—Treatment of β -7 as described for the preparation of α -8 gave β -8 (76%) as a white foam; R_f 0.11 (4:1 ether–petroleum ether); $[\alpha]_D^{27} - 26^\circ$ (c 0.67,

CHCl_3); $\nu_{\text{max}}^{\text{KBr}}$ 3490 (OH), 2956 (CH), 2869 (CH), and 1757 cm^{-1} (C=O). ^1H NMR (CDCl_3): δ 5.62 (bs, 1 H, NOH), 5.33 (m, 1 H, =CH chol), 5.20 (t, 1 H, $J_{3,2} = J_{3,4} = 10$ Hz, H-3), 5.12 (t, 1 H, $J_{4,5} = 10$ Hz, H-4), 5.05 (dd, 1 H, $J_{2,1} = 7.5$ Hz, H-2), 4.62 (d, 1 H, H-1), 4.20 (m, 2 H, H-6a,b), 3.97 (m, 1 H, H-1'a), 3.85 (m, 1 H, H-1'b), 3.73 (m, 1 H, H-5), 3.48 (m, 2 H, H-7'a,b), 3.13 (tt, 1 H, H-3 chol), 2.83 (m, 2 H, H-2'a,b), 2.65 (m, 2 H, H-4'a,b), 2.50–0.60 (m, chol skeleton), 2.10, 2.02, 2.00, and 1.98 (4 s, each 3 H, 4 OAc). EIMS: m/z (%) 831 (1.5, $\text{M}^{+} - \text{OH}$), 772 (1.5), 713 (2), 512 (10), 462 (100, $\text{M}^{+} - \text{Ochol}$), 368 (48, Ochol), 353 (20), 347 (30), 169 (92), 109 (80), 72 (100), and 55 (55). Anal. Calcd for $\text{C}_{47}\text{H}_{77}\text{NO}_{12}$ (848.14): C, 66.56; H, 9.15; N, 1.65. Found: C, 66.46; H, 9.17; N, 1.73.

3-Acetoxy-3-aza-7-(cholest-5-en-3 β -yloxy)heptyl 2,3,4,6-tetra-O-acetyl- α -D-glucopyranoside (α -9).—A solution of α -8 (0.2 g, 0.24 mmol) in pyridine (2 mL) and Ac_2O (0.5 mL) was kept at 20°C for 16 h. Usual work-up followed by column chromatography (4:1 ether–petroleum ether) afforded α -9 (0.2 g, 95%) as a white foam; R_f 0.15 (4:1 ether–petroleum ether); $[\alpha]_{\text{D}}^{23} + 45.2^\circ$ (c 1.5, CHCl_3); $\nu_{\text{max}}^{\text{KBr}}$ 2935 (CH), 2902 (CH), and 1754 cm^{-1} (C=O). ^1H NMR (CDCl_3): δ 5.48 (t, 1 H, $J_{3,2} = J_{3,4} = 10$ Hz, H-3), 5.35 (m, 1 H, =CH chol), 5.10 (t, 1 H, $J_{4,5} = 10$ Hz, H-4), 5.05 (d, 1 H, $J_{1,2} = 3.5$ Hz, H-1), 4.85 (dd, 1 H, H-2), 4.30 (m, 1 H, $J_{5,6b} = 4$, $J_{6a,6b} = 12.5$ Hz H-6b), 4.15–4.00 (m, 2 H, $J_{5,6a} = 2$ Hz, H-6a, H-5), 3.85 (m, 1 H, H-1'b), 3.65 (m, 1 H, H-1'a), 3.48 (m, 2 H, H-7'a,b), 3.12 (m, 2 H, H-2'a,b), 2.90 (m, 2 H, H-4'a,b), 2.50–0.60 (m, chol skeleton), 2.12, 2.10, 2.08, 2.03, and 2.01 (5 s each 3 H, 5 OAc). EIMS: m/z (%) 674 (1), 448 (2), 404 (10), 389 (12), 322 (100), 309 (32), 216 (43), 148 (35), 127 (55), 83 (80), 74 (98), and 60 (50). Anal. Calcd for $\text{C}_{49}\text{H}_{79}\text{NO}_{13}$ (890.17): C, 66.12; H, 8.95; N, 1.57. Found: C, 66.02; H, 8.90; N, 1.74.

3-Acetoxy-3-aza-7-(cholest-5-en-3 β -yloxy)heptyl 2,3,4,6-tetra-O-acetyl- β -D-glucopyranoside (β -9).—Obtained from β -8, as described for the preparation of α -9, in 91% yield as a white foam; R_f 0.12 (4:1 ether–petroleum ether); $[\alpha]_{\text{D}}^{20} - 21.4^\circ$ (c 1.1, CHCl_3); $\nu_{\text{max}}^{\text{KBr}}$ 2934 (CH) and 1756 cm^{-1} (C=O). ^1H NMR (CDCl_3): δ 5.35 (m, 1 H, =CH chol), 5.20 (t, 1 H, $J_{3,2} = J_{3,4} = 10$ Hz, H-3), 5.12 (t, 1 H, $J_{4,5} = 10$ Hz, H-4), 5.00 (dd, 1 H, $J_{2,1} = 7$ Hz, H-2), 4.63 (d, 1 H, H-1), 4.28 (m, 1 H, H-6b), 4.12 (m, 1 H, H-6a), 3.92 (m, 1 H, H-1'b), 3.72 (m, 2 H, H-1'a + H-5), 3.45 (m, 2 H, H-7'a,b), 3.10 (m, 3 H, H-3 chol + H-2'a,b), 2.85 (m, 2 H, H-4'a,b), 2.50–0.60 (m, chol skeleton), 2.10, 2.08, 2.06, 2.02, and 2.00 (5 s, each 3 H, 5 OAc). EIMS: m/z (%) 531 (1), 441 (3), 386 (33, Ochol), 368 (18), 301 (20), 275 (22), 157 (80), 140 (25), 98 (95), 86 (95), and 60 (100). Anal. Calcd for $\text{C}_{49}\text{H}_{79}\text{NO}_{13}$ (890.17): C, 66.12; H, 8.95; N, 1.57. Found: C, 66.38; H, 8.66; N, 1.57.

3-Aza-7-(cholest-5-en-3 β -yloxy)-3-hydroxyheptyl α -D-glucopyranoside (α -10).—A solution of α -8 (0.33 g, 0.39 mmol) in 0.02 M methanolic NaOMe was kept at 20°C for 14 h, neutralized (0.1 M aq AcOH), concentrated, then subjected to flash column chromatography (15:2:3 CHCl_3 –acetone–MeOH) to give α -10 (0.24 g, 90.3%) as a white foam; R_f 0.20 (15:2:3 CHCl_3 –acetone–MeOH); $[\alpha]_{\text{D}}^{24} + 12.4^\circ$ (c 0.74, CHCl_3); $\nu_{\text{max}}^{\text{KBr}}$ 3355 (OH), 2935 (CH), and 2848 cm^{-1} (CH). ^1H NMR (pyridine- d_5 , 60°C): δ 7.80 (s, 1 H, N-OH), 5.40 (m, 1 H, =CH chol), 5.25 (d, 1 H, $J_{1,2} = 3.8$ Hz, H-1), 4.65–4.40 (m, 3 H, H-2,3,4), 4.30 (m, 2 H, H-1'a,b), 4.20–3.90 (m, 3 H, H-5 + H-6a,b), 3.45 (t, 2 H, H-7'a,b), 3.20 (m, 1 H, CH chol), 3.10 (t, 2 H, H-2'a,b), 2.83 (t, 2 H,

H-4'a,b), and 2.50–0.50 (m, cholesterol + 2 CH₂). EIMS: m/z (%) 633 (0.5), 470 (17), 386 (32, Ochol), 308 (100), 353 (28), 294 (43, M⁺ – Ochol), 247 (32), 145 (70), 95 (85), 81 (90), 73 (98), and 55 (100). Anal. Calcd for C₃₉H₆₉NO₈ (679.99): C, 68.89; H, 10.23; N, 2.06. Found: C, 68.60; H, 10.31; N, 2.15.

3-Aza-7-(cholest-5-en-3 β -yloxy)-3-hydroxyheptyl β -D-glucopyranoside (β -10).—Obtained from β -8, as described for the preparation of α -10, in 88% yield as a white foam; R_f 0.22 (15:2:3 CHCl₃–acetone–MeOH); $[\alpha]_D^{20}$ –24.3° (c 1.2, CHCl₃); ν_{\max}^{KBr} 3355 (OH), 2945 (CH), and 2836 cm^{–1} (CH). ¹H NMR (pyridine-*d*₅): δ 7.80 (bs, 1 H, N-OH), 5.38 (m, 1 H, =CH chol), 4.75 (d, 1 H, $J_{1,2}$ 7.5 Hz, H-1), 4.60–3.95 (m, 7 H, $J_{5,6b}$ 2.5, $J_{6a,6b}$ 12, $J_{5,6a}$ 5.5 Hz, H-6a,b + H-1'a,b + H-2,3,4), 3.87 (m, 1 H, $J_{4,5}$ 9 Hz, H-5), 3.48 (t, 2 H, H-7'a,b), 3.15 (m, 3 H, H-2'a,b + CH chol), 2.83 (t, 2 H, H-4'a,b), 2.60–0.60 (m, cholesterol + 2 CH₂). EIMS: m/z (%) 663 (0.5, M⁺ – OH), 648 (10.6), 470 (5), 386 (20, Ochol), 368 (100), 353 (20), 294 (12, M⁺ – Ochol), 213 (20), 159 (32), 145 (42), and 90 (72). Anal. Calcd for C₃₉H₆₉NO₈ (679.99): C, 68.89; H, 10.23; N, 2.06. Found: C, 68.68; H, 10.24; N, 2.12.

4-Aza-6-ethyl-4-(2-ethylhexanoyloxy)-5-oxodecyl 2,3,4,6-tetra-O-acetyl-1-thio- β -D-glucopyranoside (12).—To a solution of **11** (1.15 g, 2.63 mmol) in a mixture of CHCl₃ (50 mL) and pyridine (5 mL) was added dropwise 2-ethylhexanoyl chloride (1.7 mL, 10.5 mmol). After 2 h at 20°C, water (2 mL) was added and the concentrated mixture subjected to column chromatography (4:1 \rightarrow 3:2 hexane–EtOAc) to give **12** (1.4 g, 77%) as a syrup; R_f 0.37 (3:2 hexane–EtOAc); $[\alpha]_D^{20}$ –25.2° (c 1.2, CHCl₃); $\lambda_{\max}^{\text{EtOH}}$ 205 nm (ϵ 5894); ν_{\max}^{NaCl} 2961 (CH), 1780 (C=O), 1757 (C=O), and 1670 cm^{–1} (C=O). ¹H NMR (CDCl₃): δ 5.10 (t, 1 H, $J_{3,2} = J_{3,4} = 9.5$ Hz, H-3), 5.07 (t, 1 H, $J_{4,5}$ 9.5 Hz, H-4), 5.00 (t, 1 H, $J_{2,1}$ 9.5 Hz, H-2), 4.50 (d, 1 H, H-1), 4.23 (m, 1 H, $J_{5,6b}$ 5, $J_{6a,6b}$ 12.5 Hz, H-6b), 4.10 (m, 1 H, $J_{5,6a}$ 3 Hz, H-6a), 3.90–3.60 (m, 3 H, H-5 + 2 CH), 2.90–1.10 (m, alkyl chains), 2.10, 2.08, 2.02, and 1.98 (4 s, each 3 H, 4 OAc), and 1.10–0.70 (4 t, each 3 H, 4 Me). EIMS: m/z (%) 629 (0.6, M⁺ – AcOH), 531 (1), 509 (0.6), 331 (20), 215 (12), 169 (38), 127 [22, COCH₂Et(CH₂)₃Me], 109 (18), 99 (28), 88 (67), and 57 (100). Anal. Calcd for C₃₃H₅₅NO₁₂S (689.87): C, 57.40; H, 8.04; N, 2.03; S, 4.65. Found: C, 57.60; H, 8.11; N, 2.06; S, 4.87.

4-Aza-6-ethyl-4-hydroxy-5-oxodecyl 1-thio- β -D-glucopyranoside (13).—A solution of **12** (0.5 g, 0.72 mmol) in 0.02 M methanolic NaOMe (50 mL) was kept 16 h at 24°C, then neutralized (0.1 M aq AcOH). The mixture was concentrated and subjected to column chromatography (9:1 EtOAc–MeOH) to give **13** (0.23 g, 80%); mp 59.1–59.8°C; R_f 0.14 (9:1 EtOAc–MeOH); $[\alpha]_D^{23}$ –36° (c 1.0, CHCl₃); $\lambda_{\max}^{\text{EtOH}}$ 204 nm (ϵ 281); ν_{\max}^{KBr} 3374 (OH), 2930 (CH), and 1606 cm^{–1} (C=O). ¹H NMR (CDCl₃): δ 9.55 (bs, 1 H, N-OH), 5.10 (d, 1 H, OH), 5.00 (d, 1 H, OH), 4.92 (d, 1 H, OH), 4.47 (t, 1 H, $J_{6a,b,OH}$ 6 Hz, CH₂OH), 4.12 (d, 1 H, $J_{1,2}$ 9.5 Hz, H-1), 3.70–1.00 (m, 19 H, H-2,3,4,5,6a,b, CH, and 6 CH₂), and 1.00–0.70 (2 t, each 3 H, 2 Me). EIMS: m/z (%) 396 (0.2, M⁺ + 1), 359 (0.9), 312 (0.4), 275 (0.2), 233 (0.8), 145 (5), 127 [17, COCH₂Et(CH₂)₃Me], 99 (11), 88 (36), 73 (58), and 57 (100). Anal. Calcd for C₁₇H₃₃NO₇S · 0.5 H₂O (404.52): C, 50.47; H, 8.47; N, 3.46; S, 7.93. Found: C, 50.53; H, 8.35; N, 3.50; S, 7.87.

2-(2,3,4,6-Tetra-O-acetyl- α -D-glucopyranosyl)ethanal (15).—Compound **14** (1 g, 2.7 mmol) was treated with OsO₄ and NaIO₄ as described for the preparation of α -2. Column chromatography (7:3 hexane–acetone) yielded **15** (0.75 g, 75%) as white

crystals; mp 71.5–72.5°C; R_f 0.14 (7:3 hexane–acetone); ν_{\max}^{KBr} 2952 (CH), 2825 (CH), 2727 (CH), and 1742 cm^{-1} (C=O). ^1H NMR (CDCl_3): δ 9.72 (t, 1 H, $J_{2',1'a,b}$ 2 Hz, H-2'), 5.26 (t, 1 H, $J_{3,2} = J_{3,4} = 8$ Hz, H-3), 5.15 (dd, 1 H, $J_{2,1}$ 5.5 Hz, H-2), 5.00 (t, 1 H, $J_{4,5}$ 8 Hz, H-4), 4.88 (dt, 1 H, $J_{1,1'a,b}$ 7.5 Hz, H-1), 4.25 (m, 1 H, $J_{5,6b}$ 5.5, $J_{6a,6b}$ 12.5 Hz, H-6b), 4.07 (m, 1 H, $J_{5,6a}$ 3 Hz, H-6a), 3.88 (m, 1 H, H-5), 2.80 (m, 2 H, H-1'a,b), and 2.25–2.90 (4 s, each 3 H, 4 OAc). EIMS: m/z (%) 331 (0.4, $\text{M}^{++} - \text{Ac}$), 315 (1, $\text{M}^{++} - \text{OAc}$), 301 (1), 212 (22), 169 (52), 139 (100), 115 (21), 97 (40), and 69 (13). Anal. Calcd for $\text{C}_{16}\text{H}_{22}\text{O}_{10}$ (374.35): C, 51.34; H, 5.92. Found: C, 50.77; H, 5.90.

(E)- and (Z)-1-(Hydroxyimino)-2-(2,3,4,6-tetra-O-acetyl- α -D-glucopyranosyl)ethane (16).—Compound 15 (1.4 g, 3.74 mmol) treated as described for the preparation of α -3 yielded, after column chromatography (7:3 hexane–acetone), 16 (1.2 g, 82%) as white crystals; mp 149–149.5°C; R_f 0.11 (7:3 hexane–acetone); $[\alpha]_{\text{D}}^{22} + 63^\circ$ (c 1.8, CHCl_3); ν_{\max}^{KBr} 3250 (OH), 3045 (CH), 2950 (CH), and 1743 cm^{-1} (C=O). ^1H NMR (CDCl_3): δ 7.78 (bs, 1 H, N-OH), 6.78 (dd, 1 H, $J_{2',1'a}$ 4.5, $J_{2',1'b}$ 6 Hz, H-2'), 5.35 (t, 1 H, $J_{3,4} = J_{3,2} = 9$ Hz, H-3), 5.12 (dd, 1 H, $J_{2,1}$ 5.5 Hz, H-2), 5.00 (t, 1 H, $J_{4,5}$ 9 Hz, H-4), 4.46 (dt, 1 H, $J_{1,1'a,b}$ 6.5 Hz, H-1), 4.25 (m, 1 H, $J_{5,6b}$ 5, $J_{6a,6b}$ 13 Hz, H-6b), 4.10 (m, 1 H, $J_{5,6a}$ 3 Hz, H-6a), 3.93 (m, 1 H, H-4), 3.10 (ddd, 1 H, $J_{1'a,1'b}$ 16 Hz, H-1'b), 2.50 (dt, 1 H, H-1'a), and 2.12–1.98 (4 s, each 3 H, 4 OAc). EIMS: m/z (%) 390 (2, $\text{M}^{++} + 1$), 331 (1.5, $\text{M}^{++} - \text{OAc}$), 312 (0.5), 270 (2), 227 (2), 211 (4), 169 (74), 154 (29), 127 (32), 109 (100), 97 (38), 81 (41), 70 (39), 61 (20), and 55 (19). Anal. Calcd for $\text{C}_{46}\text{H}_{23}\text{NO}_{10}$ (389.36): C, 49.36; H, 5.95; N, 3.60. Found: C, 48.96; H, 5.79; N, 3.71.

N-Hydroxy-2-(2,3,4,6-tetra-O-acetyl- α -D-glucopyranosyl)ethylamine (17).—Compound 16 (1 g, 2.57 mmol) was treated with NaBH_4 as described for the preparation of α -4, to yield 17 (0.67 g, 67%) as white crystals; mp 116.5–118.0°C; R_f 0.39 (19:1 CHCl_3 –MeOH); $[\alpha]_{\text{D}}^{25} + 63.6^\circ$ (c 0.9, CHCl_3); ν_{\max}^{KBr} 3477 (NH), 3230 (OH), 2938 (CH), 1739 (C=O), and 1720 cm^{-1} (C=O). ^1H NMR (CDCl_3): δ 6.30–5.70 (bs, 2 H, NHOH), 5.31 (t, 1 H, $J_{3,2} = J_{3,4} = 9$ Hz, H-3), 5.07 (dd, 1 H, $J_{2,1}$ 4 Hz, H-2), 4.97 (t, 1 H, $J_{4,5}$ 9 Hz, H-4), 4.30 (m, 1 H, H-1), 4.25 (m, 1 H, $J_{5,6b}$ 5.5, $J_{6a,6b}$ 12 Hz, H-6b), 4.10 (m, 1 H, $J_{5,6a}$ 3 Hz, H-6a), 3.95 (m, 1 H, H-5), 3.05 (t, 2 H, H-2'a,b), 2.40–1.90 (4 s, each 3 H, 4 OAc), \sim 2.20 (m, 1 H, H-1'b), and 1.70 (m, 1 H, H-1'a). EIMS: m/z (%) 392 (4.6, $\text{M}^{++} + 1$), 350 (0.6), 331 (0.9), 289 (1), 272 (1.2), 212 (4.6), 169 (24), 137 (33), 124 (78), 97 (26), 81 (40), and 46 (100). Anal. Calcd for $\text{C}_{16}\text{H}_{25}\text{NO}_{10}$ (391.38): C, 49.10; H, 6.44; N, 3.58. Found: C, 49.10; H, 6.45; N, 3.66.

2-Ethyl-N-(2-ethylhexanoyloxy)-N-[2-(2,3,4,6-tetra-O-acetyl- α -D-glucopyranosyl)ethyl]hexanamide (18).—To a solution of 17 (0.3 g, 0.77 mmol) in a mixture of CH_2Cl_2 (20 mL) and pyridine (1 mL) was added dropwise 2-ethylhexanoyl chloride (0.28 mL, 1.72 mmol). After 2 h at 20°C, water (2 mL) was added and the concentrated mixture subjected to column chromatography (7:3 hexane–EtOAc) to give 18 (0.57 g, 91%) as a syrup; R_f 0.22 (7:3 hexane–EtOAc); $[\alpha]_{\text{D}}^{20} + 47^\circ$ (c 0.9, CHCl_3); ν_{\max}^{NaCl} 2961 (CH), 2874 (CH), 1781 (C=O), 1752 (C=O), and 1670 cm^{-1} (C=O). ^1H NMR (CDCl_3): δ 5.30 (t, 1 H, $J_{2,3} = J_{3,4} = 9$ Hz, H-3), 5.10 (dd, 1 H, $J_{2,1}$ 5.5 Hz, H-2), 4.98 (t, 1 H, $J_{4,5}$ 9 Hz, H-4), 4.23 (m, 2 H, $J_{5,6b}$ 5, $J_{6a,6b}$ 12.5 Hz, H-6b + H-1), 4.10 (m, 1 H, $J_{5,6a}$ 3 Hz, H-6a), 3.90 (m, 2 H, H-5 + H-2'b), 3.60 (m, 1 H, H-2'a), 2.40 (m, 2 H, 2 CH), 2.20–1.10 (m, alkyl chain), 2.12, 2.10, 2.08, and 1.98 (4 s, each 3 H, 4 OAc), and 1.10–0.75 (4 t, each 3 H, 4 Me). EIMS: m/z (%) 644 (0.5, M^{++}), 531 (0.4),

442 (10), 397 (2), 264 (10), 127 [60, COCH₂(CH₂)₃Me], 99 (48), and 57 (100). Anal. Calcd for C₃₂H₅₃NO₁₂ (643.78): C, 59.70; H, 8.30; N, 2.18. Found: C, 59.53; H, 8.28; N, 2.22.

2-Ethyl-N-(2-ethylhexanoyloxy)-N-[2-(α -D-glucopyranosyl)ethyl]hexanamide (19).—A solution of **18** (0.5 g, 0.78 mmol) in 0.02 M methanolic NaOMe (50 mL) was kept 4 h at 20°C, then neutralized (0.1 M aq AcOH). The mixture was concentrated and subjected to column chromatography (15:2:3 CHCl₃–acetone–MeOH) to give **19** (0.12 g, 31%) and **20** (0.17 g, 60%) as syrups; *R_f* 0.55 (15:2:3 CHCl₃–acetone–MeOH); [α]_D²² +41.3° (c 0.6, CHCl₃); $\nu_{\text{max}}^{\text{KBr}}$ 3390 (OH), 2960 (CH), 1765 (C=O), and 1644 cm⁻¹ (C=O). ¹H NMR (CDCl₃, 55°C): δ 4.08 (dt, 1 H, *J*_{1,2} 5, *J*_{1,1'a,b} 9.5 Hz, H-1), 3.90–3.60 (m, 10 H, H-2,3,4,5,6a,b + 4 OH), 2.40 (m, 2 H, 2 CH), 2.15–1.10 (m, alkyl chain), and 1.10–0.75 (4 t, each 3 H, 4 Me). EIMS: *m/z* (%) 476 (1, M⁺), 458 (2, M⁺ – OH), 440 (1.2), 423 (1.5), 404 (1.8), 349 [1.5, M⁺ – COCH(Et)(CH₂)₃Me], 331 (1.8), 127 [23, COCH(Et)(CH₂)₃Me], 99 [32, Me(CH₂)₃CH₂Et], and 57 [100, Me(CH₂)₃]. Anal. Calcd for C₂₄H₄₅NO₈ · 0.5 H₂O (484.63): C, 59.48; H, 9.56; N, 2.89. Found: C, 59.76; H, 9.63; N, 2.92.

2-Ethyl-N-[2-(α -D-glucopyranosyl)ethyl]-N-hydroxyhexanamide (20).—Obtained as described for the preparation of **19**; *R_f* 0.33 (15:2:3 CHCl₃–acetone–MeOH); [α]_D²¹ +120° (c 0.4, CHCl₃); $\nu_{\text{max}}^{\text{KBr}}$ 3353 (OH), 2960 (CH), 2867 (CH), and 1605 cm⁻¹ (C=O). ¹H NMR (CD₃COCD₃): δ 8.80 (bs, 1 H, N-OH), 4.40–4.10 (m, 3 H, 3 × OH), 3.95 (dt, 1 H, H-1), 3.85–3.10 (m, 7 H, H-2,3,4,5,6a,6b,CH₂OH), 3.02 (m, 1 H, CH), 1.98 (m, 2 H, H-2'a,b), 1.70–1.10 (m, alkyl chain), and 1.00–0.80 (2 t, each 3 H, 2 Me). EIMS: *m/z* (%) 348 (0.5, M⁺ – 1), 331 (1, M⁺ – H₂O), 296 (2.5), 277 (5), 262 (2), 144 (10), 127 [15, COCH(Et)(CH₂)₃Me], 99 [22, CH(Et)(CH₂)₃Me], 88 (12), and 57 [100, Me(CH₂)₃]. Anal. Calcd for C₁₆H₃₁NO₇ · 0.5 H₂O (358.43): C, 53.61; H, 9.00; N, 3.91. Found: C, 53.81; H, 8.94; N, 3.94.

References

- [1] R. Miethchen, H. Prade, J. Holz, K. Praefcke, and D. Blunk, *Chem. Ber.*, 126 (1993) 1707–1712.
- [2] C. Baron and T.E. Thompson, *Biochim. Biophys. Acta*, 382 (1975) 276–285.
- [3] R.A. Schwendener, M. Asanger, and H.G. Weder, *Biochem. Biophys. Res. Commun.*, 100 (1981) 1055–1062.
- [4] O. Lockhoff, *Angew. Chem., Int. Ed. Engl.*, 30 (1991) 1611–1620.
- [5] Y. Osa, E. Kaji, K. Takahashi, M. Hirooka, S. Zen, and F.W. Lichtenthaler, *Chem. Lett.*, (1993) 1567–1570.
- [6] W. Spevak, J.O. Nagy, D.H. Charych, M.E. Schaefer, J.H. Gilbert, and M.D. Bednarski, *J. Am. Chem. Soc.*, 115 (1993) 1146–1147.
- [7] D.M. Gordon and S.J. Danishefsky, *J. Am. Chem. Soc.*, 114 (1992) 659–663.
- [8] J.M.J. Tronchet, in R.I. Zhdanov (Ed.), *Bioactive Spin Labels*, Springer, Berlin, 1992, pp 355–387, and references therein.
- [9] J.M.J. Tronchet, M. Zsély, O. Lassout, I. Komaromi, M. Geoffroy, and R.A. Schwendener, VIIth Eur. Carbohydr. Symp., Cracow, Poland, 1993, Abstr. A-036.
- [10] R.T. Lee and Y.C. Lee, *Carbohydr. Res.*, 37 (1974) 193–201.
- [11] E.A. Talley, M.D. Vale, and E. Yanovsky, *J. Am. Chem. Soc.*, 67 (1945) 2037–2039.
- [12] R.F. Borch, M.D. Bernstein, and H.D. Durst, *J. Am. Chem. Soc.*, 93 (1971) 2897–2904.

- [13] J.M.J. Tronchet, G. Zosimo-Landolfo, N. Bizzozero, D. Cabrini, F. Habashi, E. Jean, and M. Geoffroy, *J. Carbohydr. Chem.*, 7 (1988) 169–186.
- [14] J.M.J. Tronchet, M. Zsély, A. Ricca, F. Barbalat-Rey, and M. Geoffroy, *Biochimie*, 74 (1992) 57–62.
- [15] J.M.J. Tronchet, E. Winter-Mihaly, J. Rupp, F. Barbalat-Rey, and M. Geoffroy, *Carbohydr. Res.*, 136 (1985) 375–390.
- [16] D. Horton and T. Miyaka, *Carbohydr. Res.*, 184 (1988) 221–229.
- [17] A. Ricca, J.M.J. Tronchet, J. Weber, and Y. Ellinger, *J. Phys. Chem.*, 96 (1992) 10779–10784.
- [18] J.M.J. Tronchet, N. Bizzozero, M. Koufaki, F. Habashi, and M. Geoffroy, *J. Chem. Res.*, (1989) (S) 334, (M) 2601–2620.
- [19] B.S. Furniss, A.Z. Hannaford, P.W.G. Smith, and A.R. Tatchell, *Vogel's Textbook of Practical Chemistry*, 5th ed., Longman, London, 1989, pp 692–693.
- [20] W. Meyer zu Reckendorf, *Chem. Ber.*, 96 (1963) 2019–2021.
- [21] W.C. Still, M. Kahn, and A. Mitra, *J. Org. Chem.*, 43 (1978) 2923–2925.
- [22] F. Barbalat-Rey and P. Lichtle, unpublished results.